

## Synthesis of a New Polycyclic Quinone by Reduction of a Dihydrobenzo[a]naphthacenequinone

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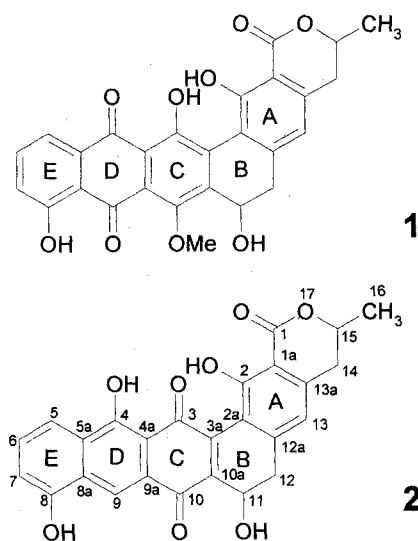
Presently, more than 20 compounds have been reported containing the dihydrobenzo[a]naphthacenequinone skeleton such as the pradimicins<sup>1)</sup>, benanomycins<sup>2)</sup>, WS79089-compounds<sup>3)</sup>, ericamycin<sup>4)</sup> and others. Previously we isolated benaphthamycin and WS79089A (**1**) from *Streptomyces* sp. HKI-0057<sup>5)</sup>. During our chemical derivatization program for naturally occurring compounds we subjected **1** to a series of reduction procedures using complex hydrides (NaBH<sub>4</sub>, NaAlH<sub>4</sub>, LiAlH<sub>4</sub>, (*t*-Butyl)<sub>2</sub>AlH) and catalytic hydrogenation (Pt/H<sub>2</sub>, Pd/H<sub>2</sub>). The initial aim was to open the lactone ring and to introduce an aldehyde or a primary alcohol structure at C-1. With the exception of NaAlH<sub>4</sub> the above reductions furnished mixture of products. However, reduction by NaAlH<sub>4</sub> in excess and subsequent hydrolysis under acidic conditions afforded a violet compound showing a lower R<sub>f</sub> on TLC than **1**. The product was purified in 30 % yield by preparative TLC, and its structure was unambiguously assigned by mass spectrometry and NMR spectroscopy (**2**; Fig. 1).

A molecular weight of 458 and the formula C<sub>26</sub>H<sub>18</sub>O<sub>8</sub> were readily suggested by HREI-MS (*m/z* 440.0908 ([M-H<sub>2</sub>O]<sup>+</sup>; calcd. 440.0913) and ESI-MS (*m/z* 457.3 [M-H]<sup>-</sup>). The <sup>1</sup>H NMR spectrum displayed signals of a methyl group (H-16, 1.45 ppm, d), two methylene (H-12, 2.86 ppm; H-14, 3.01 and 3.12 ppm) and two methine groups (H-11, 5.08 ppm; H-15, 4.85 ppm). In addition, six aromatic protons were visible. Two appeared as singlets (H-9, 8.41 ppm; H-13, 6.92 ppm). The others (H-5, 7.81 ppm; H-6, 7.55 ppm; H-7, 7.2 ppm) displayed the typical coupling pattern of three neighboured aromatic protons.

Signals at 11.0, 11.8, 13.2 and 5.22 ppm were attributable to the hydroxyl protons. The <sup>13</sup>C NMR spectrum showed 26 signals, attributable to the quinone (186.3 and 181.0 ppm) and lactone carbonyls (169.6 ppm), three phenolic (158.7, 159.6, 155.9 ppm), two aliphatic oxygen-bonded (57.5 and 76.2 ppm) and a series of aromatic and aliphatic carbons (see Experimental). For the assignment of the quaternary carbons the C, H long-range coupled NMR spectra (HMBC) were particularly helpful. Thus, the visible <sup>2</sup>J and <sup>3</sup>J<sub>C,H</sub> long-range correlations (Fig. 2) and weak <sup>4</sup>J<sub>C,H</sub> correlations between H-9 and C-2, and H-13 and C-1 supplied supporting evidence.

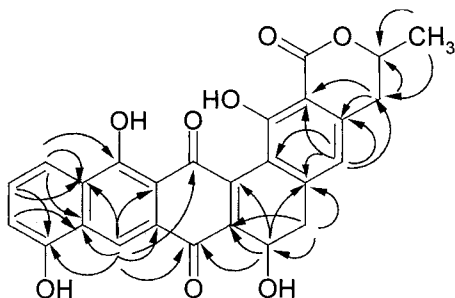
In comparison with **1** the new quinone structure **2** (2,4,8,11-tetra-hydroxy-15-methyl-11,12,14,15-tetrahydro-17-oxa-hexaphene-1,3,10-trione) harbours the quinone in the C ring instead in D ring. Moreover, during NaAlH<sub>4</sub> reduction and subsequent acidification under aerobic conditions one molecule of methanol was eliminated. The mechanism of this reductive rearrangement-elimination reaction is presently under discussion. It is remarkable that solely NaAlH<sub>4</sub> provoked the formation of **2**. Moreover, the observed rearrangement reaction was not observed when madurahydroxylactone was used as another representative of the dihydrobenzo[a]naphthacenequinones<sup>6)</sup>. Compound **2** displayed moderate activity against Gram-positive bacteria

Fig. 1. Chemical constitutions of WS79089 A (**1**) and new quinone (**2**).



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Fig. 2. Observable C, H long-range correlations in the HMBC spectrum of **2**.



such as *Bacillus subtilis* ATCC 6633 and *Staphylococcus aureus* SG511 during standardized agar diffusion assay<sup>7)</sup>. Thus 50  $\mu\text{g}$  **2** dissolved in 50  $\mu\text{l}$  methanol afforded inhibition zones with 16 mm and 13 mm diameter, respectively. No activity was found against Gram-negative bacteria and fungi. The  $\text{IC}_{50}$  values of **2** were determined with 72 hours L929, K562, and HeLa cells ( $\text{IC}_{50}$ ) to amount to 1.9, 1.7 and 5.4  $\mu\text{g}/\text{ml}$ , respectively. Comparable biological activities were found with WS79089A (**1**)<sup>3)</sup>.

### Experimental

Mass spectra were recorded with a high-resolution sector-field spectrometer AMD 402 (AMD Intectra, Bremen, Germany) and triple-quadrupole instrument Quattro equipped with an electrospray ion source (VG Biotech, Altrincham, England). 1D and 2D NMR spectra ( $^1\text{H}$ ,  $^{13}\text{C}$ ,  $^1\text{H}$ ,  $^1\text{H}$ -COSY, HSQC, NOESY, HMBC) were recorded with a 500 MHz NMR spectrometer Bruker Avance DRX 5000. Determination of cytotoxicity using the analyzer system CASY 1 was described earlier<sup>8)</sup>.

#### Synthesis of **2**

20 mg **1** dissolved in 50 ml dry tetrahydrofuran was stirred in a 100 ml three-necked vessel under nitrogen. The solution was cooled to  $-78^\circ\text{C}$ , and 50 mg  $\text{NaAlH}_4$  was added in small portions. After 5~6 hours stirring at  $-78^\circ\text{C}$  the reaction mixture was added to a concentrated aqueous solution of  $\text{NaHSO}_3$  and, thereafter, the pH was adjusted to 1.0. Subsequently the mixture was extracted with diethylether, the ether phase dried over  $\text{Na}_2\text{SO}_4$  and concentrated *in vacuo*.

The residue was chromatographed on silica gel plates

(Merck,  $\text{CHCl}_3/\text{MeOH}$ , 97:3). In addition to the starting material running with  $R_f$  0.8, a violet zone with  $R_f$  0.35 was visible. It was extracted with  $\text{CHCl}_3/\text{MeOH}$  (1:1) and the solution was evaporated. Yield: 7.5 mg (30%), violet solid, mp.  $250\sim 252^\circ\text{C}$  (decomposition).

HREI-MS:  $m/z$  440.0908 ( $[\text{M}-\text{H}_2\text{O}]^+$ , calcd. 440.0913 for  $\text{C}_{26}\text{H}_{16}\text{O}_7$ ).  $^1\text{H}$  NMR (500 MHz in  $\text{DMSO}-d_6$ ,  $\delta$  in ppm, TMS as internal standard): 1.45 (H-16, d, 7.0 Hz), 2.86 (H-12, dd, 16.0 Hz, 3.0 Hz), 3.00 (H-14<sub>A</sub>, d, 2.5 Hz), 3.12 (H-14<sub>B</sub>, d, 2.5 Hz), 4.85 (H-15, m), 5.08 (H-11, d, t, br), 7.2 (H-7, d, 8.2 Hz), 7.55 (H-6, dd, 8.3 Hz, 8.2 Hz), 7.81 (H-5, d, 8.3 Hz), 8.41 (H-9, s), 5.22 (11-OH, br), 11.0 (8-OH, s), 11.8 (2-OH), 13.2 (4-OH).  $^{13}\text{C}$  NMR (125 MHz,  $\delta$  in ppm, in  $\text{DMSO}-d_6$ , TMS as internal standard): 20.2 (C-16), 33.5 (C-14), 37.1 (C-12), 57.8 (C-11), 76.2 (C-15), 107.2 (C-13a), 109.0 (C-4a), 114.3 (C-5), 114.4 (C-7), 115.1 (C-9), 116.0 (C-2a), 119.5 (C-13), 125.9 (C-9a), 125.9 (C-8a), 128.5 (C-5a), 130.5 (C-6), 140.7 (C-3a), 142.8 (C-1a), 144.6 (C-10a), 147.0 (C-12a), 155.9 (C-9), 158.7 (C-2), 159.6 (C-4), 169.6 (C-1), 181.0 (C-10), 186.3 (C-3).

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